Corrections

Orientational Distribution of α-Helices in the Colicin B and El Channel Domains: A One and Two Dimensional ^{15}N Solid-State NMR Investigation in Uniaxially Aligned Phospholipid Bilayers, byStephan Lambotte, Pieter Jasperse, and Burkhard Bechinger*, Volume 37, Number 1, January 6, 1998, pages 16−22.

Page 18. In column 1, the sentence beginning on line 50 should read as follows: A total of 65 rows were recorded in t_1 with a dwell time of 39.4 μs.

Page 19. In order to demonstrate that a significant proportion of the protein is oriented within the NMR sample, the spectrum in Figure 4 is represented with the contour lines drawn at low level. Therefore, not all intensities represent protein resonances. The number of resolved peaks can only be determined accurately once further technical developments become available such as methods for the assignment of resonances.

Page 21. The following should be included under Acknowledgment: We would like to take this opportunity to extend our grateful thanks to the members of the Resource for Solid-State NMR of Proteins at the University of Pennsylvania (Grant P41RR09731 from the Biomedical Research Technology Program, National Center for Research Resources, National Institutes of Health). In particular, we would like to express again our thanks to Dr. Kathleen Valentine for discussions, introduction into the goals and instrumentation of the Resource, and her help with the 700 MHz spectrometer. Our thanks also include Dr. Ron McNamara and Tai Van Le, who helped in providing a working infrastructure. We apologize that a detailed acknowledgment was not made previously.

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Crystal Structure of Fragment Double-D from Human Fibrin with Two Different Bound Ligands, byStephen J. Everse, Glen Spraggon, Leela Veerapandian, Marcia Riley, and Russell F. Doolittle*, Volume 37, Number 24, June 16, 1998, pages 8637−8642.

Page 8638. The fourth sentence in column 1 should read as follows: Protein solutions containing 10 mg of fragment double-D/mL of 50 mM Tris buffer, pH 7.0, 5 mM $CaCl_2$, 10 mM GPRPam, and 10 mM GHRPam were mixed in equal volume (5 μL + 5 μL) with well solution containing 50 mM Tris, pH 8.0, 20 mM $CaCl_2$, 12% PEG 3350, and 2 mM sodium azide.

Page 8641. In column 2, the sentence beginning on line 44 should read as follows: It should be noted that the preparations used in our X-ray study were crystallized in the presence of 12.5 mM calcium.

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Pigment Epithelium-Derived Factor (PEDF) Binds to Glycosaminoglycans: Analysis of the Binding Site, by Elena Alberdi, C. Craig Hyde, and S. Patricia Becerra*, Volume 37, Number 30, July 28, 1998, pages 10643−10652.

Page 10649. In the legend to Figure 8, the accession number for bovine PEDF (bPEDF) should read U48229.

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MgATP-Dependent and MgATP-Independent [^{3}H]Noradrenaline Release from Perforated Synaptosomes Both Use *N*-Ethylmaleimide-Sensitive Fusion Protein, byXu Zheng and Joseph A. Bobich*, Volume 37, Number 36, September 8, 1998, pages 12569−12575.

Page 12575. In column 1, ref *27* should read ref *29* on line 39, and ref *29* should read ref *28* on line 49.

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Kinetic Studies on the Effect of Yeast Cofilin on Yeast Actin Polymerization, byJinyan Du and Carl Frieden*, Volume 37, Number 38, September 22, 1998, pages 13276−13284.

Page 13280. Figure 5 is incorrect as printed. The figure should appear as follows:

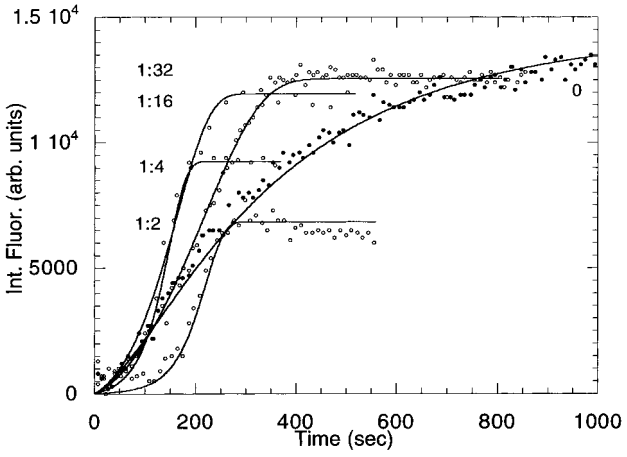

FIGURE 5: Polymerization of yeast actin (0.25 mg/mL, 5.9 μM) as a function of cofilin concentration at pH 6.6 and 20 °C. The time course of polymerization measured using intrinsic fluorescence and the reaction was initiated by the addition of 1 mM Mg^{2+} at pH 6.6 after preincubation to replace tightly bound Ca^{2+} with Mg^{2+} as described in Materials and Methods. The molar cofilin to actin ratios are given in the figure. The symbols represent data collected continuously while the solid lines are the fitted data using the parameters given in the legend to Figure 1 and in Table 1.

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Effects of L-Histidine and Its Structural Analogues on Human *N*-Myristoyltransferase Activity and Importance of EEVEH Amino Acid Sequence for Enzyme Activity, by Rajala V. S. Raju, Raju S. S. Datla, Robert C. Warrington, and Rajendra Sharma*, Volume 37, Number 42, October 20, 1998, pages 14928−14936.

Page 14929. In the legend to Table 1, line 2, "Transferase assay was carried out with WT (1.0 μg/assay), N^6 (1.2 mg/assay), N^{21} (1.2 μg/assay)" should read "Transferase assay was carried out with WT (1.0 μg/assay), N^8 (1.2 μg/assay), N^{20} (1.2 μg/assay)". In line 4, "initiated by the addition of 50 mM [1-^{14}C]myristoyl-CoA" should read "initiated by the addition of 50 μM [1-^{14}C]myristoyl-CoA".

Page 14933. In column 2, line 23, "Substitution of combinations of Glu-289 and -290, Glu-289 and -290, and Glu-290 and -292" should read "Substitution of combinations of Glu-289 and -290, Glu-289 and -292, and Glu-290 and -292".

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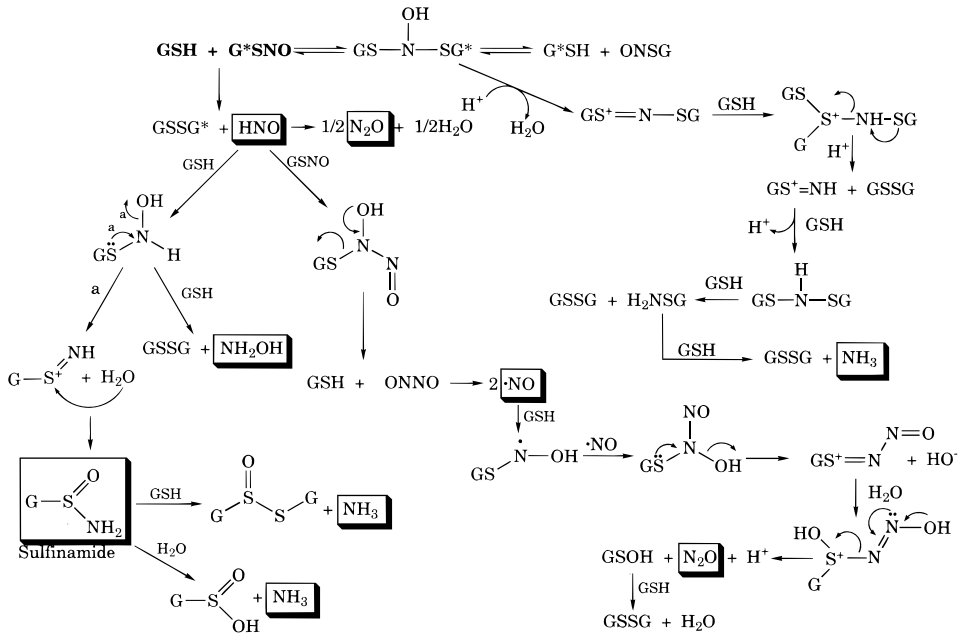
Reaction between *S*-Nitrosothiols and Thiols: Generation of Nitroxyl (HNO) and Subsequent Chemistry, by Patrick S.-Y. Wong, Jinjoo Hyun, Jon M. Fukuto,* Frances N. Shirota, Eugene G. DeMaster, Don W. Shoeman, and Herbert T. Nagasawa, Volume 37, Number 16, April 21, 1998, pages 5362−5371.

Page 5369. Scheme 3 has a slight error regarding the mechanism by which ammonia is generated. The scheme should appear as follows:



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